

## An arsenal of naturopathic compounds for the treatment of COVID-19: A comprehensive review

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### Abstract

Despite the tremendous efforts to find a reasonable potent and efficacious drug candidate for the treatment of COVID-19, there is not a magic drug to cure the symptoms of this deadly disease. This study aims to give some information about natural compounds against SARS-CoV-2 and their ethnopharmacology, and the possible antiviral mechanism of especially plant secondary metabolites. Nowadays, although several NCEs were approved for the treatment of COVID-19 or to ease the symptoms, most of these new medicines seem to eventually be less effective in large populations. Moreover, it is imperative to use prophylactic agents and natural remedies to prevent infections as well as boost the immune system. When we look at the epidemiological and geographical movement of viral infections, it is obvious that the countries that follow naturopathic/homeopathic treatment options seem to recover faster than the other countries. Thus, it is extremely important to identify what natural treatment recipe works for certain households. For methodology, Web of Science, Sci-Finder, Google Scholar, Science Direct, Scopus, TUBIVES, CITES, and other databases were used as initial origins to look for related articles and apply search terms. In this review, many medicinal and aromatic plants, natural products, and commercial compounds were screened against SARS-CoV-2. Furthermore, it gives details about these compounds and related medicinal and aromatic plants. Also, this comprehensive review sheds light on current natural treatment options reported in the literature and the future projections to halt this deadly disease via natural medicines.

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Natural products, COVID-19 treatment, SARS-CoV-2 activity, antiviral phytochemicals, phytotherapy agents, homeopathic medicine.

## 1. Introduction

COVID-19 was seen for the first time in the Hubei Province of the People's Republic of China on 30

January 2019, World Health Organization Emergency Committee declared a global health emergency,



briefly some studies determined that viral particles can enter the human body by respiratory tract [1] and the patients with COVID-19 have symptoms such as fever, shortness of breath, dry cough [2], dysgeusia and anosmia [3]. Also, this virus can penetrate the ACE-2 receptor found in some organs such as lungs and kidneys [4], it affects not only the respiratory system but also the gastrointestinal, nervous, and hepatic systems [5]. After this virus isolated and identified, some antiviral commercial drug as remdesivir [6], favipiravir [7], chloroquine [8], and natural products (Xiu Powder, An Gong Niu Huang Pill, Hou Po Xia Ling Decoction) utilized in treatment of COVID-19 [9]. Natural active compounds have significant properties for lung health, such as reducing lung damage, preventing the development of pneumonia, suppressing the emission of proteins on the alveolar surface, and minimizing lung damage [10]. Almost one million metabolites are estimated to be produced throughout the plant kingdom, and most of them have not been discovered, yet [11]. Nowadays, natural compounds were used in the treatment of some diseases in the 20 plant families such as Arecaceae, Aristolochiaceae, Cucurbitaceae, Piperaceae, Rubiaceae, Ranunculaceae, Zingiberaceae in a totally 450 families [12]. Especially for COVID-19, many natural compounds and their antiviral mechanisms were reported. One study reported that bioflavonoids from *Torreya nucifera* negatively affected SARS-CoV 3CLpro's replication [13]. Moreover, the combination of 13 herbs (such as Forsythia suspense (Thunb.), *Ephedra sinica* Stapf Cao, and *Gypsum fibrosum*) not only suppressed the replication of SARS-CoV-2 but also led to a change in the structure of virus cells [14]. For using these active compounds in the treatment of many diseases, these plant-based compounds have many advantages as safe for human consumption, not environmental pollution, and more acceptable to pharmaceuticals and local farmers [15, 16]. In view of the fact that these recent studies, this study aims to give some information about natural compounds against SARS-CoV-2 and their ethnopharmacology.

## 2. Materials and methods

This review consists of many parts such as ethnopharmacology of natural products for COVID-

19 treatment, important natural products for COVID-19 treatment, plant primary and secondary metabolites and mainly antiviral mechanisms. All the databases were collected by electronic libraries which are Web of Science, SciFinder, Google Scholar, Science Direct and Scopus. Also, most of the details were given about plants from many book chapters and published books. The scientific names of medicinal and aromatic plants were checked by using Natural Agriculture Library in the U.S. Department of Agriculture (USDA), Medicinal Plant Services in Royal Botanic Garden, Türkiye's Plants Data Service (TÜBİVES), the Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES). All the primary and secondary metabolites found in Dr. Duke's Phytochemical and Ethnobotanical Databases in USDA and their chemical structures were checked by PubChem.

## 3. Results and discussion

### 3.1. Ethnopharmacology of natural products for COVID-19 treatment

Plants used ethnobotanically in the treatment of flu and colds were used in the symptomatic treatment of coronavirus disease during the pandemic period. It is noteworthy that the plants selected for the studies against the coronavirus were selected from those with antiviral effect potential and used in the treatment of influenza [17, 18]. For this reason, many plants are used ethnobotanically in the treatment of flu and colds listed in this study. Looking at different ethnobotanical studies, *R. canina* stands out as the most used plant against flu and colds. According to one study's results, volunteers and patients with flu took medication with 2 g of liquid rosehip and placebo for six months during the winter, respectively. After six months, the incidence of symptoms of the flu and cold (coughing, headache, muscle stiffness, and fatigue) decreased statistically in patients compared to volunteers [19].

Following rosehip, *A. sativum*, *U. dioica*, *S. nigra*, *R. officinalis*, *O. vulgare*, *M. spicata* and *M. longifolia*, *Eucalyptus* species are among other frequently used plants. Each of them draws attention to the plants whose effects are discussed during the COVID-19 pandemic. It can be considered as having the potential to act of most plants against coronavirus. The fact that

these plants, briefly summarized in Table 1 and some of them given in Fig. 1 (drawn by Biorender), were compiled from ethnobotanical information is an indication that there is also reliable human use.



**Figure 1.** Some medicinal and aromatic plants

### 3.2. Important natural products for COVID-19 treatment

The COVID-19 pandemic, still being present since 2019, created awareness about human immunology health. The scientific community has made significant progress in mitigating the threat of COVID-19 through the discovery and development of vaccines which can be classified as Inactivated and Protein subunit, Viral vector, and mRNA vaccines. On the contrary to common belief about most of plant-based vaccine leading to allergenic conditions [20], The Novavax, saponin-based Matrix M adjuvant, showed great efficacy against COVID-19 of 89% [21]. One of plant-based vaccine is the NVX-CoV2373 (Novavax) vaccine, developed by using the bark of the *Quillaja saponaria* (Soapbark) tree, and was efficacious in preventing Sars-CoV-2- B.1.351 variant. A total of 4387 participants (2199 with NVX-CoV2373 and 2188 with placebo) were randomized and dosed at least once. The vaccine efficacy was 49.4% and 60.1% in 2684 baseline seronegative participants and HIV-negative symptomatic of COVID-19, respectively. Noticeably, in the post-hoc analysis, the vaccine efficacy was 51.0% in HIV-negative participants and 43.0% in the combined HIV-negative and PLWH population [22]. Another example is the CoVLP+AS03 vaccine, which showed anti-viral activity against SARS-CoV-2 by efficacy ranging from 69.5% against symptomatic infection to 78.8% against moderate-to severe disease. (Funded by Medicago; ClinicalTrials.gov number, NCT04636697.) [23].

Not only vaccines but merely monoclonal antibodies utilized in treatment of the disease. Naturally immune serum and panels of monoclonal antibodies used against the Omicron variants known as COVID-19 sublineages. The virus had reduced neutralization by serum from a person vaccinated with 3 doses of Pfizer or AstraZeneca vaccine rather than BA 1 and BA 2 variants [24]. However, promising drugs do not exist. Thus, scientists reported that the secret to a healthy life is to use functional nutrients for a powerful immune system, sleep quality and exercise. On the other hand, some of these lines for protecting against this disease are isolation, using vitamins, and avoiding contact with another person. Particularly, the separate property of COVID-19 from other viral diseases is an asymptomatic course of the disease, and using some antiviral products based on medicinal and aromatic compounds can prevent this problem. However, there was no evidence found any of these products using clinical. Although it suggests that using aromatherapeutic products decreases stress in human life during the pandemic, it is exactly known that these products can be harmful such as lung irritation by using unconscious, so we must know some documents for using right these products. "Pharmacopoeia" is used as a document for making medicine in Ancient Greek, and these documents defined as involving physicochemical properties, some qualitative and quantitative methods of inactive and active compounds in pharmaceutical production, and rules to follow at both national and international levels [25]. On the other hand, monographs are described as active and inactive compounds of one pharmacophore like chemical/ biological, plant-derived medicines, comments, and properties (physical, soluble, boiling, and melting temperatures). Thanks to these analyses for achieving character analysis of completed product's dosage, keeping storage conditions and impurity. Furthermore, the quality control analyses of these products contain some analytical methods such as stability, microbiological purity, and solubility [26]. Shed light on these traditional medicines [27], and small molecule agents [28, 29], this scenario, bioactivenatural products [30, 31] playing an irreplaceable role in the treatment of SARS-CoV-2 infection.

**Table 1.** Plants used ethnobotanically for flu and common cold

Plant name	Usage	Reference
<i>Achillea vermicularis</i> Trin.	Int. Dec.	[60]
<i>Aconitum heterophyllum</i> Wall. ex Royle.	Int. Dec.	[61]
<i>Acorus calamus</i> L.	Int. Inf.	[62]
<i>Adhatoda vasica</i> Nees.	Int. Inf.	[63]
<i>Allium sativum</i> L.	Int.	[64]
<i>Arbutus unedo</i> L.	Int. Dec.	[65]
<i>Artemisia afra</i> Jacq.ex Willd.	Int. Inf.	[66]
<i>Artemisia macrocephala</i> Jacquem. ex Besser	Int. Inf.	[61]
<i>Artemisia scoparia</i> Waldst. et Kit	Int. Dec.	[67]
<i>Capsicum annuum</i> L.	Int. Dec.	[61]
<i>Conium sphaerocarpum</i> Hilliard & B.L.	Ext. Inf.	[66]
<i>Conyza scabrida</i> DC	Int. Inf.	[66]
<i>Crocus sativus</i>	Int. Inf.	[68]
<i>Cydonia oblonga</i> Mill.	Int. Dec.	[69]
<i>Cymbopogon citratus</i>	Ext. Inf.	[70]
<i>Dactylorhiza osmanica</i> (Klinge) P.F. Hunt & Summerh.	Int. Dec.	[65]
<i>Ephedra gerardiana</i> Wall. exStapf ex Decne.	Int. Inf.	[61]
<i>Eucalyptus camaldulensis</i> Dehnhardt.	Int. Dec.	[61]
<i>Eucalyptus globulus</i> L.	Int. Dec.	[64]
<i>Hedera helix</i> L.	Int. Dec.	[71]
<i>Juniperus drupacea</i> Labill.	Int. Dec.	[65]
<i>Lavandula dentata</i>	Inf.Dec.	[72]
Table 1. Continued	Fum.	
<i>Leonotis nepetifolia</i>	Int. Inf.	[70]
<i>Lepidium sativum</i> L.	Int. Inf.	[71]
<i>Mentha pulegium</i>	Int. Dec.	[64]
<i>Mentha spicata</i> L.	Int. Inf.	[73]
<i>Nepeta cataria</i>	Int. Inf.	[66]
<i>Nigella sativa</i> L.	Int.	[74]
<i>Orchis anatolica</i> Boiss.	Int. Dec.	[65]
<i>Pistacia terebinthus</i> L.	Int. Dec.	[74]
<i>Plantago ovata</i> Forssk.	Int. Dec.	[61]
<i>Punica granatum</i> L.	Int. Dec.	[61]
<i>Rosa canina</i> L.	Int. Dec.	[74]
<i>Rosmarinus officinalis</i> L.	Int. Inf.	[64]
<i>Salvia moorcroftiana</i> Wall. ex Benth	Int. Powder	[61]
<i>Thymus kotschyanus</i> Boiss. & Hohen.	Int. Inf.	[73]

### 3.2.1. Whole natural plants products

Recently, it is known that medicinal and aromatic plants are used for the development of some nutrients against many diseases [32-35]. Especially, COVID-19 [36, 37]. In one of the studies, we emphasize *Artemisia annua* may be better candidate to prophylactically control SARS-COV-2 infection [38]. In addition, in Swiss, the clinical experiments of artemi C oral spray product, curcumin and artemisinin, are still ongoing for treatment in SARS-COV-2 infection [39].

### 3.2.2. Isolated/elucidated natural products

These plant natural compounds are classified as primary and secondary metabolites according to their role and biosynthetic pathway [40], in addition, the secondary metabolites are produced by only one species or related groups, whereas primary metabolites are produced from all plant kingdoms [41].

#### 3.2.2.1. Plant primary metabolites for the treatment of COVID-19

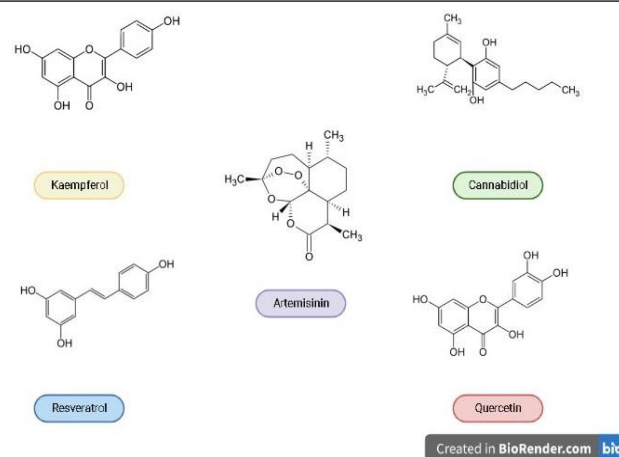


The plant primary metabolism used for surviving, growing, enveloping, and communicating with nature [42], and they produced by the shikimate pathway, TCA cycle, glucose pathway [43] which their precursor molecules are CO<sub>2</sub>, H<sub>2</sub>O, and NH<sub>3</sub> in these pathways [44]. One of the primary metabolites is hesperidin obtained from *Citrus* sp. [45] prevents virus replication via binding main protein receptors of SARS-CoV-2 such as spike glycoprotein and protease domain. By virtue of it binds to ACE-2 receptors, virus cannot hold on, this molecule can be utilized for the treatment of COVID-19 [46].

#### 3.2.2.2. Plant secondary metabolites of natural products for the treatment of COVID-19

Under some abiotic (drought, salty, elevated temperature, UV radiation) and biotic conditions (some microorganisms, insects, and nematodes in the soil), plants produce some secondary metabolites for defense and adapting the nature. These secondary metabolites are utilized by Mevalonic acid, Methylerythritol and shikimate pathways and shikimic acid, the amino acids phenylalanine/tyrosine, tryptophan, ornithine, and lysine are precursor molecules in these pathways [47,48]. These metabolites consist of the main groups such as phenolics, terpenes, and nitrogen-containing compounds [48]. Focus on the COVID-19, some secondary metabolites such as emodin, palmatine are suggested as potential drug candidates against SARS-CoV-2 [49, 50] (Table 2), [51-54]. Quercetine increased caspase activity [55], kaemferol inhibited TNF- $\alpha$ , IL-6, IL-1 $\beta$  cytokinins and increase SOD [56], resveratrol inhibited MAPK, PI3K, AKT [57], and also cannabidiol inhibited expression of TNF- $\alpha$  and IL-6 cytokines [58], (Fig. 2, drawn by Biorender).

Particularly, these natural compounds have some roles in antiviral disease which has cancellation to penetration to the host cell (lectin), adhesion to lungs and reproduction (alkaloids) of coronavirus, as well as inhibition of binding to the surface of host cells by changing proteins in the surface of virus cells (polyphenols), prevent to replication of virus by inhibition reverse transcriptase enzyme (flavonoids and terpenes), block translation proteins and recognize viral proteins for replication, repress active caspase 1 for preventing cytokine storm [59].



**Figure 2.** The structures of plant secondary metabolite

## 4. Conclusion and future perspectives

Natural treatment will always be center of the attention since the regulations and applications as well as availability are easier than the drugs. The main advantage of the use of natural medicines are cost-effectiveness since the new COVID-19 medicines will be expensive and non-affordable for some countries. Thus, naturopathic treatment options will always be a popular alternative. Because of the exponentially growing food supplement market in the last decades, effective and less costly natural medicines as well as research and development around this concept will be one the most attractive areas for researchers who study natural products. Additionally, in the future more medicines will be translated from natural compounds to potentially treat certain therapeutic areas including infectious diseases. In conclusion, several natural products and commercial compounds were screened against SARS CoV-2. Among those screened compounds decent antiviral activity has been observed. Some of these natural compounds and their analogs demonstrated reasonable antiviral activity against SARS CoV-2. In future, some of these compounds are worth investigating their anti-viral activity in vivo models. Natural medicines will always be the center of attention for the treatment of viral and bacterial infections because of the availability and cost-effectiveness of these supplements compared to expensive and not easily accessible drugs.

## Abbreviations

3CLpro: 3-chymotrypsin like protease; ACE: Mitogen-activated protein kinase; AKT: Activation protein

**Table 2.** The Possible Antiviral Mechanism of Plant Secondary Metabolites

Secondary metabolite	Biological activity mechanism	References
<b>Alkaloids</b>		
Cepharantine	Inhibit MPro	[75]
Emetine	Inhibit RdRp	[76]
Hyoscyamine	Inhibit S protein	[77]
Incanumine	Inhibit 3C-like protease	[78]
Magnoflorine	Inhibit S protein	[52]
Neoechinulin A	Inhibit MPro	[79]
Nigellidine	Inhibit ACE-1 receptor	[80]
<b>Flavonoids</b>		
Afzelin	Inhibit MPro, hACE-2 and RdRp	
Amentoflavone	Inhibit ACE-2 receptor	[82]
Baicalein	Inhibit RdRp	[83]
Biflavone	Inhibit PLpro	[84]
Chrysin	Inhibit S protein and ACE-2 receptor	[85]
Hesperidin	Inhibit to bind ACE-2 receptor	[86]
Luteolin	Inhibit ACE-2 receptor	[87]
Quercetin 3-glucuronide-7-glucoside	Inhibit MPro and ACE-2 receptor	[53]
<b>Phenolic compounds</b>		
Deoxyshikonin	Inhibit MPro	[88]
Corilagin	Exhibit SARS-CoV-2-RBD and hACE2 protein	[89]
Coumarine	Inhibit 3CLpro	[90]
ellagic acid	Inhibit RBD-ACE-2 receptor	[91]
Epicatechin	Inhibit ACE-2 receptor	[92]
Kazinol J	Inhibit PLpro	[93]
Resveratrol	Inhibit MAPK, PI3K, AKT	[54]
Shikonin	Inhibit MPro	[94]
Theaflavin	Inhibit RdRp	[95]
<b>Terpenes</b>		
Abietane	Inhibit PLpro	[96]
Cannabidiol	Inhibit expression of TNF- $\alpha$ and IL-6 cytokines	[97]
Carvacrol	Inhibit ACE-2 receptor	[52]
Nimocin	Inhibit M and E proteins	[98]
Parthenolide	Reduce IL-1, IL-2, IL-6, IL-8, and TNF- $\alpha$ production pathways	[99]
Rosmariquinone	Inhibit PLpro	[100]
Saikosaponin C	Inhibit NSP15 endoribonuclease	[101]

activated protein kinase; AKT: Activation protein kinase B; CAT: Catalase; CITES: Convention on International Trade in Endangered Species of Wild Fauna and Flora; COVID-19: Coronavirus disease-19; COX-2 Cyclooxygenase-2; Dec: Decoction; Ext: External; IFN- $\gamma$ : Interferon gamma; IL: Interleukin; Inf: Infusion; Int: Internal; MAPK: Mitogen-activated protein kinase; MPro: Main protease; NF- $\kappa$ B: Nuclear factor-kappa B; Nsp15: Non-structural protein 15; PI3K: Phosphoinositide 3-kinase; PLpro: papain-like protease; RBD: Receptor binding domain; RdRp: RNA-dependent; RNA: polymerase; S: Spike; SARS-

CoV: Anti-severe acute respiratory syndrome coronavirus; SOD: Superoxide dismutase; TNF- $\alpha$ : Tumor necrosis factor- $\alpha$ ; TUBIVES: Türkiye's Plants Data Service; TNF- $\alpha$ : Tumor necrosis factor- $\alpha$ ; USDA: U.S. Department of Agriculture; WHO: World Health Organization.

### Authors' contributions

Designed this study, M.G.; Extracted all necessary data for this review, A.E.K., M.T., K.F.E., Drawn all figures, M.T.; Revised this study, M.G., M.T.

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## Availability of data and materials

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## Conflicts of interest

The authors declare no conflict of interest.

## References

1. Leung, N.H.L.; Chu, D.K.W.; Shiu, E.Y.C.; Chan, K.-H.; McDevitt, J.J.; Hau, B.J.P.; Yen, H.-L.; Li, Y.; Ip, D.K.M.; Peiris, J.S.M.; Seto, W.-H.; Leung, G.M.; Milton, D.K.; Cowling, B.J. Respiratory virus shedding in exhaled breath and efficacy of face masks. *Nat. Med.* 2020, 26, 676–680. <https://doi.org/10.1038/s41591-020-0843-2>.
2. Chen, H.; Guo, J.; Wang, C.; Luo, F.; Yu, X.; Zhang, W.; Li, J.; Zhao, D.; Xu, D.; Gong, Q.; Liao, J.; Yang, H.; Hou, W.; Zhang, Y. Clinical characteristics and intrauterine vertical transmission potential of COVID-19 infection in nine pregnant women: a retrospective review of medical records. *Lancet.* 2020, 395, 809–815. [https://doi.org/10.1016/S0140-6736\(20\)30360-3](https://doi.org/10.1016/S0140-6736(20)30360-3).
3. Fan, C.; Lei, D.; Fang, C.; Li, C.; Wang, M.; Liu, Y.; Bao, Y.; Sun, Y.; Huang, J.; Guo, Y.; Yu, Y.; Wang, S. Perinatal transmission of 2019 coronavirus disease—associated severe acute respiratory syndrome coronavirus 2: Should we worry?. *Clin. Infect. Dis.* 2021, 72, 862–864. <https://doi.org/10.1093/cid/ciaa226>.
4. Turner, A.J.; Hiscox, J.A.; Hooper, N.M. ACE2: from vasopeptidase to SARS virus receptor. *Trends Pharmacol. Sci.* 2004, 25, 291–294. <https://doi.org/10.1016/j.tips.2004.04.001>.
5. Chen, Y.; Liu, Q.; Guo, D. Emerging coronaviruses: Genome structure, replication, and pathogenesis. *J. Med. Virol.* 2020, 92, 418–423. <https://doi.org/10.1002/jmv.25681>.
6. Wang, M.; Cao, R.; Zhang, L.; Yang, X.; Liu, J.; Xu, M.; Shi, Z.; Hu, Z.; Zhong, W.; Xiao, G. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res.* 2020, 30, 269–271. <https://doi.org/10.1038/s41422-020-0282-0>.
7. Yamamura, H.; Matsuura, H.; Nakagawa, J.; Fukuoka, H.; Domi, H.; Chujoh, S. Effect of favipiravir and an anti-inflammatory strategy for COVID-19. *Crit. Care.* 2020, 24, 413. <https://doi.org/10.1186/s13054-020-03137-5>.
8. Colson, P.; Rolain, J.-M.; Lagier, J.-C.; Brouqui, P.; Raoult, D. Chloroquine and hydroxychloroquine as available weapons to fight COVID-19. *Int. J. Antimicrob. Agent.* 2020, 55, 105932. <https://doi.org/10.1016/j.ijantimicag.2020.105932>.
9. Luo, L.; Jiang, J.; Wang, C.; Fitzgerald, M.; Hu, W.; Zhou, Y.; Zhang, H.; Chen, S. Analysis on herbal medicines utilized for treatment of COVID-19. *Acta Pharm. Sin. B.* 2020, 10, 1192–1204. <https://doi.org/10.1016/j.apsb.2020.05.007>.
10. Anand, A.V.; Balamuralikrishnan, B.; Kaviya, M.; Bharathi, K.; Parithathvi, A.; Arun, M.; Senthilkumar, N.; Velayuthaprabhu, S.; Saradhadevi, M.; Al-Dhabi, N.A.; Arasu, M.V.; Yatoo, M.I.; Tiwari, R.; Dhama, K. medicinal plants, phytochemicals, and herbs to combat viral pathogens including SARS-CoV-2. *Molecules.* 2021, 26, 1775. <https://doi.org/10.3390/MOLECULES26061775>.
11. Afendi, F.M.; Okada, T.; Yamazaki, M.; Hirai-Morita, A.; Nakamura, Y.; Nakamura, K.; Ikeda, S.; Takahashi, H.; Altaf-Ul-Amin, M.; Darusman, L.K.; Saito, K.; Kanaya, S. KNApSACk family databases: Integrated metabolite-plant species databases for multifaceted plant research. *Plant Cell Physiol.* 2012, 53. <https://doi.org/10.1093/PCP/PCR165>.
12. Reddy. Important Medicinal Plant Families and Plant Based Drugs: A Review, in: Eares.Org. 2017, pp. 277–283. <https://doi.org/10.17758/EAP.AE0317304>.
13. Ryu, Y. B.; Hyung J. J.; Jang Hoon K.; Young M. K.; Ji-Young P.; Doman K., Thi Thanh H. N. et al. Biflavonoids from *Torreya nucifera* displaying SARS-CoV 3CLpro inhibition. *Bioorg. Med. Chem. Lett.* 2010, 18, 22, 7940-7947. <https://doi.org/10.1016/j.bmc.2010.09.035>.
14. Runfeng, L.; Hou Y.; Huang J.; Pan W.; Ma Q.; Shi Y.; Li C. et al. Lianhuaqingwen exerts anti-viral and anti-inflammatory activity against novel coronavirus (SARS-CoV-2). *Pharmacol. Res. Commun.* 2020, 156, 104761. <https://doi.org/10.1016/j.phrs.2020.104761>.
15. Bhattacharjee, R.; Dey, U. An overview of fungal and bacterial biopesticides to control plant pathogens/diseases. *Afr. J. Microbiol. Res.* 2014, 8, 1749–1762. <https://doi.org/10.5897/AJMR2013.6356>.
16. Sogvar, O.B.; Koushesh Saba, M.; Emamifar, A. Aloe vera and ascorbic acid coatings maintain postharvest quality and reduce microbial load of strawberry fruit. *Postharvest. Biol. Technol.* 2016, 114, 29–35. <https://doi.org/10.1016/J.POSTHARVBIO.2015.11.019>.
17. Rafiqul Islam, A.T.M.; Ferdousi, J.; Shahinozzaman, M. Previously published ethno-pharmacological reports

- reveal the potentiality of plants and plant-derived products used as traditional home remedies by Bangladeshi COVID-19 patients to combat SARS-CoV-2. Saudi J. Biol. Sci. 2021, 28, 6653–6673. <https://doi.org/10.1016/J.SJBS.2021.07.036>.
18. Tegen, D.; Dessie, K.; Dامتie, D. Candidate Anti-COVID-19 Medicinal Plants from Ethiopia: A review of plants traditionally used to treat viral diseases. J. Evid. Based Complement. Altern. Med. 2021. <https://doi.org/10.1155/2021/6622410>.
  19. Winther, K.; Warholm, L.; Campbell-Tofte, J.; Marstrand, K. Effect of *Rosa canina* L.(Rose-hip) on cold during winter season in a middle-class population: A randomized, double-blinded, placebo-controlled trial. J. Herb Med. 2018, 13, 34-41. <https://doi.org/10.1016/j.hermed.2018.04.003>.
  20. Guan, Z.J.; Guo, B.; Huo, Y.L.; Guan, Z.P.; Dai, J.K. Wei, Y.H. Recent advances and safety issues of transgenic plant-derived vaccines. Appl. Microbiol. Biotechnol. 2013, 97, 2817-2840. <https://doi.org/10.1007/s00253-012-4566-2>.
  21. Shinde, V.; Sutika B.; Zaheer H.; Moherndran A.; Qasim B.; Lee F.; Umesh L. et al. Efficacy of NVX-CoV2373 Covid-19 vaccine against the B. 1.351 variant." N. Engl. J. Med. 2021. 384, 20, 1899-1909. <https://doi.org/10.1056/NEJMoa2103055>.
  22. Hager, Karen J.; Gonzalo Pérez, M.; Philipe G.; Ricardo S. D.; Gretchen H.; Conrado L.; Alexander I. M. et al. Efficacy and safety of a recombinant plant-based adjuvanted Covid-19 vaccine. New Engl. J. Med. 2022, 386, 22, 2084-2096. <https://doi.org/10.1056/NEJMoa2201300>.
  23. Creech, C.B.; Walker, S.C.; Samuels, R.J.SARS-CoV-2 vaccines. JAMA. 2021, 325,13, 1318-1320. <https://doi.org/10.1001/jama.2021.3199>.
  24. Tuekprakhon, A.; Nutalai, R.; Djokaite-Guraliuc, A.; Zhou, D.; Ginn, H.M.; Selvaraj, M.; Liu, C.; Mentzer, A.J.; Supasa, P.; Duyvesteyn, H.M. Das, R. Antibody escape of SARS-CoV-2 Omicron BA. 4 and BA. 5 from vaccine and BA. 1 serum. Cell. 2022, 185, 14, 2422-2433. <https://doi.org/10.1016/j.cell.2022.06.005>.
  25. European Directorate for the Quality of Medicines.Supplements . European Pharmacopoeia 9.8.2022.
  26. European medicines agency. Herbal Medicinal Products. 2022. <https://www.edqm.eu/en/>.
  27. Wang, Z. And Yang, L.Chinese herbal medicine: Fighting SARS-CoV-2 infection on all fronts. J. Ethnopharmacol. 2021, 270, 113869. <https://doi.org/10.1016/j.jep.2021.113869>.
  28. Wang, Z.; Yang, L. and Song, X.Q., Oral GS-441524 derivatives: Next-generation inhibitors of SARS-CoV-2 RNA-dependent RNA polymerase. Front. Immunol. 2022a. 13. <https://doi.org/10.3389/fimmu.2022.1015355>.
  29. Kolarič, A.; Jukič, M. and Bren, U. Novel small-molecule inhibitors of the SARS-CoV-2 spike protein binding to neuropilin 1. Pharmaceuticals.2022, 15, 2, 165. <https://doi.org/10.3390/ph1502016>.
  30. Yang, L. and Wang, Z. Natural products, alone or in combination with FDA-approved drugs, to treat COVID-19 and lung cancer. Biomedicines, 2021, 9, 6, 689. <https://doi.org/10.3390/biomedicines9060689>.
  31. Wang, Z.; Wang, N.; Yang, L. and Song, X.Q.. Bioactive natural products in COVID-19 therapy. Front. Pharmacol. 2022b, 13. <https://doi.org/10.3389/fphar.2022.926507>.
  32. Tepe, M.; Abadan, Ş.; Sağlam, M.F.; Süzerer, V.; Erçin, P.B.; Atilla, D.; Baykal, E.E.; Şeker, M.G.; Yağcı, T. and Çiftçi, Y.Ö. In vitro mass production, chemical modification, and cytotoxicity of shikonin derivatives on breast cancer cells. Ind. Crop. Prod. 2023, 192, 116087. <https://doi.org/10.1016/j.indcrop.2022.116087>.
  33. Karadağ, A.E.; Çaşkurlu, A.; Demirci, B.; Demirci, F. Binary synergistic combinations of Lavender and Fennel essential oils with amoxicillin. Planta Med. 2022. <https://doi.org/10.1055/A-1891-1119>.
  34. Karadağ, A.E.; İpekçi, E.; Yağcılar, A.P.; Demirbolat, İ., Kartal, M.; Siafaka, P.I.; Okur, N.Ü. Antibacterial evaluation of *Elettaria cardamomum* (L.) Maton, *Lavandula angustifolia* mill. and *Salvia fruticosa* mill. essential oil combinations in mouthwash preparations. Nat. Vol. Essen. Oil. 2020, 7, 9–17. <https://doi.org/10.37929/NVEO.685474>.
  35. Sakallı, E.A.; Teralı, K.; Karadağ, A.E.; Biltekin, S.N.; Koşar, M.; Demirci, B.; Başer, K.H.C.; Demirci, F. *In vitro* and *in silico* evaluation of ACE2 and LOX inhibitory activity of eucalyptus essential oils, 1,8-cineole, and citronellal. Nat. Prod. Commun. 2022. <https://doi.org/10.1177/1934578X221109409>.
  36. Biltekin, S.N.; Karadağ, A.E.; Demirci, B.; Demirci, F. ACE2 and LOX enzyme inhibitions of different lavender essential oils and major components linalool and camphor. ACS Omega. 2022, 7, 36561–36566. <https://doi.org/10.1021/acsomega.2c04518>.
  37. Demirci, F.; Teralı, K.; Karadağ, A.E.; Biltekin, S.N.; Ak Sakallı, E.; Demirci, B.; Koşar, M.; Başer, K.H.C. *In Vitro* and *In Silico* evaluation of ACE2 and LOX inhibitory activity of origanum essential oils and carvacrol. Planta Med. 2022. <https://doi.org/10.1055/a-1828-2479>.
  38. Dogan, K.; Erol, E.; Didem Orhan, M.; Degirmenci, Z.; Kan, T.; Gungor, A.; Yasa, B.; Avsar, T.; Cetin, Y.; Durdagi, S.; Guzel, M. Instant determination of the artemisinin from various *Artemisia annua* L. extracts by LC-ESI-MS/MS and their in-silico modelling and in



- vitro antiviral activity studies against SARS-CoV-2. *Phytochem. Anal.* 2022, 33, 303–319. <https://doi.org/10.1002/PCA.3088>.
39. Hellou, E.; Mohsin, J.; Elemy, A.; Hakim, F.; Mustafa-Hellou, M.; Hamoud, S. Effect of ArtemiC in patients with COVID-19: A Phase II prospective study. *J. Cell Mol. Med.* 2022, 26, 3281–3289. <https://doi.org/10.1111/JCMM.17337>.
  40. Böttger, A.; Vothknecht, U.; Bolle, C.; Wolf, A. Plant secondary metabolites and their general function in plants. 2018, 3–17. [https://doi.org/10.1007/978-3-319-99546-5\\_1](https://doi.org/10.1007/978-3-319-99546-5_1).
  41. Balandrin, M.F.; Klocke, J.A.; Wurtele, E.S.; Bollinger, Wm.H. Natural plant chemicals: Sources of industrial and medicinal materials. *Science*. 1985, 228, 1154–1160. <https://doi.org/10.1126/science.3890182>.
  42. Fernie, A.R.; Pichersky, E. Focus issue on metabolism: Metabolites, metabolites everywhere. *Plant Physiol.* 2015, 169, 1421–1423. <https://doi.org/10.1104/PP.15.01499>.
  43. Kroymann, J. Natural diversity and adaptation in plant secondary metabolism. *Curr. Opin. Plant Biol.* 2011, 14, 246–251. <https://doi.org/10.1016/J.PBI.2011.03.021>.
  44. Wu, S.; Chappell, J. Metabolic engineering of natural products in plants; tools of the trade and challenges for the future. *Curr. Opin. Biotechnol.* 2008, 19, 145–152. <https://doi.org/10.1016/J.COPBIO.2008.02.007>.
  45. Zhou, P.; Zheng, M.; Li, X.; Zhou, J.; Shang, Y.; Li, Z.S.; Qu, L.,. A consecutive extraction of pectin and hesperidin from *Citrus aurantium* L.: Process optimization, extract mechanism, characterization, and bio-activity analysis. *Ind. Crops. Prod.* 2022, 82, 114849. <https://doi.org/10.1016/J.INDCROP.2022.114849>.
  46. Utomo, R.Y.; Ikawati, M.; Meiyanto, E. Revealing the potency of citrus and galangal constituents to halt SARS-CoV-2. *Infection*. 2020. <https://doi.org/10.20944/PREPRINTS202003.0214.V1>.
  47. Fett-Neto, A.G.; Aoyagi, H.; Tanaka, H.; DiCosmo, F. Antitumor agents: taxol and taxanes—production by yew cell culture. *Encyclopedia of Molecular Cell Biology and Molecular Medicine*. 2006. <https://doi.org/10.1002/3527600906.MCB.200300127>.
  48. Hartmann, T. From waste products to ecochemicals: Fifty years research of plant secondary metabolism. *Phytochem.* 2007, 68, 2831–2846. <https://doi.org/10.1016/J.PHYTOCHEM.2007.09.017>.
  49. Ho, T.Y.; Wu, S.L.; Chen, J.C.; Li, C.C.; Hsiang, C.Y. Emodin blocks the SARS coronavirus spike protein and angiotensin-converting enzyme 2 interaction. *Antiviral Res.* 2007, 74, 92–101. <https://doi.org/10.1016/J.ANTIVIRAL.2006.04.014>.
  50. Jadhav, V. Anti-SARS-CoV-2 main protease complex (M pro) activity of Palmatine. Preprint (Research Square). 2020, 1–19. <https://doi.org/10.21203/rs.3.rs-38145/v1>
  51. Khazdair, M. R.; Anaeigoudari, A.; Agbor, G. A. Anti-viral and anti-inflammatory effects of kaempferol and quercetin and COVID-2019: A scoping review. *Asian Pac. J. Trop. Biomed.* 2021, 11, 8, 327–334. <https://doi.org/10.4103/2221-1691.319567>.
  52. Kiran, G.; Karthik, L.; Shree Devi, M.S.; Sathiyarajeswaran, P.; Kanakavalli, K.; Kumar, K.M.; Ramesh Kumar, D. In silico computational screening of Kabasura Kudineer - Official Siddha Formulation and JACOM against SARS-CoV-2 spike protein. *J. Ayurveda. Integr. Med.* 2022, 13, 100324. <https://doi.org/10.1016/J.JAIM.2020.05.009>.
  53. Joshi, R.S.; Jagdale, S.S.; Bansode, S.B.; Shankar, S.S.; Tellis, M.B.; Pandya, V.K.; Chugh, A.; Giri, A.P.; Kulkarni, M.J. Discovery of potential multi-target-directed ligands by targeting host-specific SARS-CoV-2 structurally conserved main protease. *J. Biomol. Struct. Dyn.* 2020, 1–16. <https://doi.org/10.1080/07391102.2020.1760137>.
  54. Giordo, R.; Zinellu, A.; Ali, H.E.; Pintus, G. Therapeutic potential of resveratrol in COVID-19-associated hemostatic disorders. *Molecules*. 2021, 856 26. <https://doi.org/10.3390/MOLECULES26040856>.
  55. Chiow, K.; Phoon, M.; Putti, T.; Tan, B. K.; Chow, V. T. Evaluation of antiviral activities of *Houttuynia cordata* Thunb. extract, quercetin, quercetrin and cinanserin on murine coronavirus and dengue virus infection. *Asian Pac. J. Trop. Med.* 2016, 9 (1), 1–7. <https://doi:10.1016/j.apjtm.2015.12.002>.
  56. Schwarz, S.; Sauter, D.; Wang, K.; Zhang, R.; Sun, B.; Karioti, A., et al. Kaempferol derivatives as antiviral drugs against the 3a channel protein of coronavirus. *Planta Med.* 2014, 80, 2/3, 177–182. <http://doi:10.1055/s-0033-1360277>.
  57. Wahedi, H. M.; Ahmad, S.; Abbasi, S. W. Stilbene-based natural compounds as promising drug candidates against COVID-19. *J. Biomol. Struct. Dyn.* 2020, 1–10. <https://doi.10.1080/07391102.2020.1762743>.
  58. Kiran G.; Karthik L.; Shree Devi M.S.; Sathiyarajeswaran P.; Kanakavalli K.; Kumar K.M.; et al. In silico computational screening of Kabasura Kudineer - official Siddha Formulation and JACOM against SARS-CoV-2 spike protein. *J. Ayurveda Integr. Med.* 2022, 13, 1. <https://doi:10.1016/j.jaim.2020.05.009>.
  59. Pal, S.; Chowdhury, T.; Paria, K.; Manna, S.; Parveen, S.; Singh, M.; Sharma, P.; Islam, S.S.; Abu Imam Saadi, S.M.; Mandal, S.M. Brief survey on phytochemicals to prevent COVID-19. *J. Indian Chem. Soc.* 2022, 99, 100244. <https://doi.org/10.1016/J.JICS.2021.100244>.

60. Mükemre, M.; Behçet, L.; Çakılciolu, U. Ethnobotanical study on medicinal plants in villages of Çatak (Van-Turkey). *J. Ethnopharmacol.* 2015, 166, 361–374. <https://doi.org/10.1016/J.JEP.2015.03.040>.
61. Kayani, S.; Ahmad, M.; Zafar, M.; Sultana, S.; Khan, M.P.Z.; Ashraf, M.A.; Hussain, J.; Yaseen, G. Ethnobotanical uses of medicinal plants for respiratory disorders among the inhabitants of Gallies – Abbottabad, Northern Pakistan. *J. Ethnopharmacol.* 2014, 156, 47–60. <https://doi.org/10.1016/J.JEP.2014.08.005>.
62. Mhlongo, L.S.; Van Wyk, B.E. Zulu medicinal ethnobotany: new records from the Amandawe area of KwaZulu-Natal, South Africa. *South Afr. J. Bot.* 2019, 122, 266–290. <https://doi.org/10.1016/J.SAJB.2019.02.012>.
63. Kichu, M.; Malewska, T.; Akter, K.; Imchen, I.; Harrington, D.; Kohen, J.; Vemulapad, S.R.; Jamie, J. F. An ethnobotanical study of medicinal plants of Chungtia village, Nagaland, India. *J. Ethnopharmacol.* 2015, 166, 5–17. <https://doi.org/10.1016/J.JEP.2015.02.053>.
64. Bouasla, A.; Bouasla, I. Ethnobotanical survey of medicinal plants in northeastern of Algeria. *Phytomedicine.* 2017, 36, 68–81. <https://doi.org/10.1016/J.PHYMED.2017.09.007>.
65. Sargin, S.A. Ethnobotanical survey of medicinal plants in Bozyazı district of Mersin, Turkey. *J. Ethnopharmacol.* 2015, 173, 105–126. <https://doi.org/10.1016/J.JEP.2015.07.009>.
66. Philander, L. An ethnobotany of Western Cape Rasta bush medicine. *J. Ethnopharmacol.* 2011, 138, 578–594. <https://doi.org/10.1016/J.JEP.2011.10.004>.
67. Au, D.T.; Wu, J.; Jiang, Z.; Chen, H.; Lu, G.; Zhao, Z. Ethnobotanical study of medicinal plants used by Hakka in Guangdong, China. *J. Ethnopharmacol.* 2008, 117, 41–50. <https://doi.org/10.1016/J.JEP.2008.01.016>.
68. Singab, A.N.B.; Ayoub, I.M.; El-Shazly, M.; Korinek, M.; Wu, T.Y.; Cheng, Y.; Bin, Chang; F.R., Wu, Y.C. Shedding the light on Iridaceae: Ethnobotany, phytochemistry and biological activity. *Ind. Crops. Prod.* 2016, 92, 308–335. <https://doi.org/10.1016/J.INDCROP.2016.07.040>.
69. Sargin, S.A.; Akçicek, E.; Selvi, S. An ethnobotanical study of medicinal plants used by the local people of Alaşehir (Manisa) in Turkey. *J. Ethnopharmacol.* 2013, 150, 860–874. <https://doi.org/10.1016/J.JEP.2013.09.040>.
70. Clement, Y.N.; Baksh-Comeau, Y.S.; Seaforth, C.E. An ethnobotanical survey of medicinal plants in Trinidad. *J. Ethnobiol. Ethnomed.* 2015, 11, 1–28. <https://doi.org/10.1186/S13002-015-0052-0/FIGURES/8>.
71. Ugulu, I.; Baslar, S.; Yorek, N.; Dogan, Y. The investigation and quantitative ethnobotanical evaluation of medicinal plants used around Izmir province, Turkey. *J. Med. Plants Res.* 2009, 3, 345–367. <https://doi.org/10.5897/JMPR.9001216>.
72. Najem, M.; Nassiri, L.; Ibibjijen, J. Vernacular names of plants between diversity and potential risks of confusion: Case of toxic plants used in medication in the central Middle Atlas, Morocco. *J. Pharm. Pharmacogn. Res.* 2021, 9, 2, 222–250.
73. Polat, R. Ethnobotanical study on medicinal plants in Bingöl (City center) (Turkey). *J. Herb Med.* 2019, 16, 100211. <https://doi.org/10.1016/J.HERMED.2018.01.007>.
74. Güzel, Y.; Güzelşemme, M.; Miski, M. Ethnobotany of medicinal plants used in Antakya: A multicultural district in Hatay Province of Turkey. *J. Ethnopharmacol.* 2015, 174, 118–152. <https://doi.org/10.1016/J.JEP.2015.07.042>.
75. Celik, S.; Akyuz, S.; Ozel, A.E. Vibrational spectroscopic characterization and structural investigations of Cepharanthine, a natural alkaloid. *J. Mol. Struct.* 2022, 1258, 132693. <https://doi.org/10.1016/j.molstruc.2022.132693>.
76. Ren, P.X.; Shang, W.J.; Yin, W.C.; Ge, H.; Wang, L.; Zhang, X.L.; Li, B.Q.; Li, H.L.; Xu, Y.C.; Xu, E.H. and Jiang, H.L. A multi-targeting drug design strategy for identifying potent anti-SARS-CoV-2 inhibitors. *Acta Pharmacol. Sin.* 2021, 43, 483–493. <https://doi.org/10.1038/s41401-021-00668-7>.
77. Skariyachan, S.; Gopal, D.; Muddebihalkar, A.G.; Uttarkar, A.; Niranjana, V. Structural insights on the interaction potential of natural leads against major protein targets of SARS-CoV-2: Molecular modelling, docking and dynamic simulation studies. *Comput Biol Med.* 2021, 132, 104325. <https://doi.org/10.1016/j.combiomed.2021.104325>.
78. Mahmood, R.A.; Hasan, A.; Rahmatullah, M.; Paul, A.K.; Jahan, R.; Jannat, K.; Bondhon, T.A.; Mahboob, T.; Nissapatorn, V.; de Lourdes Pereira, M.; Paul, T.K.; Rumi, O.H.; Wiart, C.; Wilairatana, P. Solanaceae family phytochemicals as inhibitors of 3C-like protease of SARS-CoV-2: An in silico analysis. *Molecules.* 2022, 27, 4739. <https://doi.org/10.3390/molecules27154739>.
79. Alhadrami, H.A.; Burgio, G.; Thissera, B.; Orfali, R.; Jiffri, S.E.; Yaseen, M.; Sayed, A.M.; Rateb, M.E.; Neoechinulin A as a Promising SARS-CoV-2 Mpro Inhibitor: In vitro and in silico study showing the ability of simulations in discerning active from inactive enzyme inhibitors. *Marine Drug.* 2022, 20, 163. <https://doi.org/10.3390/md20030163>.
80. Maiti, S.; Banerjee, A.; Kanwar, M. In silico Nigellidine (N. sativa) bind to viral spike/active-sites of ACE1/2, AT1/2 to prevent COVID-19 induced vasotumult/vascular-damage/comorbidity. *Vascul*

- Pharmacol. 2021, 138, 106856. <https://doi.org/10.1016/j.vph.2021.106856>.
81. Joshi T.; Joshi T.; Sharma, P.; Mathpal, S.; Pundir H.; Bhatt V.; Chandra. S. In silico screening of natural compounds against COVID-19 by targeting Mpro and ACE2 using molecular docking drug discovery from plants View Project Global Initiatives For Waste Reduction And Cutting Food Loss View Project. 2020b. [https://doi.org/10.26355/eurev\\_202004\\_21036](https://doi.org/10.26355/eurev_202004_21036).
  82. Chen, G.Y.; Pan, Y.C.; Wu, T.Y.; Yao, T.Y.; Wang, W.J.; Shen, W.J.; Ahmed, A.; Chan, S.T.; Tang, C.H.; Huang, W.C.; Hung, M.C.; Yang, J.C.; Wu, Y.C. Potential natural products that target the SARS-CoV-2 spike protein identified by structure-based virtual screening, isothermal titration calorimetry and lentivirus particles pseudotyped (Vpp) infection assay. J. Tradit. Complement. Med. 2022, 12, 73–89. <https://doi.org/10.1016/J.JTCME.2021.09.002>.
  83. Song, J.; Zhang, L.; Xu, Y.; Yang, D.; Yang, Shiyang; Zhang, W.; Wang, J.; Tian, S.; Yang, Shengqian; Yuan, T.; Liu, A.; Lv, Q.; Li, F.; Liu, H.; Hou, B.; Peng, X.; Lu, Y.; Du, G. The comprehensive study on the therapeutic effects of baicalein for the treatment of COVID-19 in vivo and in vitro. Biochem. Pharmacol. 2021, 183, 114302. <https://doi.org/10.1016/J.BCP.2020.114302>.
  84. Li, L.; Ma, L.; Hu, Y.; Li, X.; Yu, M.; Shang, H.; Zou, Z. Natural biflavones are potent inhibitors against SARS-CoV-2 papain-like protease. Phytochemistry. 2022, 193, 112984. <https://doi.org/10.1016/J.PHYTOCHEM.2021.112984>.
  85. Bachevski, D.; Damevska, K.; Simeonovski, V.; Dimova, M. Back to the basics: Propolis and COVID-19. Dermatol. Ther. 2020, 33, e13780. <https://doi.org/10.1111/DTH.13780>.
  86. Bellavite, P.; Donzelli, A. Hesperidin and SARS-CoV-2: New light on the healthy function of citrus fruits. Antioxidants. 2020, 9, 742. <https://doi.org/10.3390/ANTIOX9080742>.
  87. Ansari, W.A.; Ahamad, T.; Khan, M.A.; Khan, Z.A.; Khan, M.F. Exploration of luteolin as potential anti-COVID-19 Agent: Molecular docking, molecular dynamic simulation, ADMET and DFT Analysis. Lett. Drug. Des. Discov. 2021, 19. <https://doi.org/10.2174/1570180819666211222151725>.
  88. Kılınç, N. Naphthoquinones from onosma: molecular mechanisms of action in the treatment and prevention of COVID-19. Caucasian J. Sci. 2021, 8, 173–184. <https://doi.org/10.48138/CJO.1037727>.
  89. Yang, L.J.; Chen, R.H.; Hamdoun, S.; Coghi, P.; Ng, J.P.L.; Zhang, D.W.; Guo, X.; Xia, C.; Law, B.Y.K.; Wong, V.K.W. Corilagin prevents SARS-CoV-2 infection by targeting RBD-ACE2 binding. Phytomed. 2021, 87, 153591. <https://doi.org/10.3390/biomedicines9060689>.
  90. Abdizadeh, R.; Hadizadeh, F.; Abdizadeh, T. In silico analysis and identification of antiviral coumarin derivatives against 3-chymotrypsin-like main protease of the novel coronavirus SARS-CoV-2. Mol. Divers. 2022, 26, 1053–1076. <https://doi.org/10.1007/S11030-021-10230-6/FIGURES/15>.
  91. David, A. ben; Diamant, E.; Dor, E.; Barnea, A.; Natan, N.; Levin, L.; Chapman, S.; Mimran, L.C.; Epstein, E.; Zichel, R.; Torgeman, A. Identification of SARS-CoV-2 receptor binding inhibitors by in vitro screening of drug libraries. Molecules. 2021, 26. <https://doi.org/10.3390/MOLECULES26113213>.
  92. Al-Shuhaib, M.B.S.; Hashim, H.O.; Al-Shuhaib, J.M.B. Epicatechin is a promising novel inhibitor of SARS-CoV-2 entry by disrupting interactions between angiotensin-converting enzyme type 2 and the viral receptor binding domain: A computational/simulation study. Comput. Biol. Med. 2022, 141, 105155. <https://doi.org/10.1016/J.COMPBIOMED.2021.105155>.
  93. Choudhury, S.; Moulick, D.; Borah, A.; Saikia, P.; Mazumder, M.K. In search of drugs to alleviate suppression of the host's innate immune responses against SARS-CoV-2 using a molecular modeling approach. In Silico Pharmacol. 2021, 9, 1–9. <https://doi.org/10.1007/S40203-021-00085Y/FIGURES/3>.
  94. Jin, Z.; Du, X.; Xu, Y.; Deng, Y.; Liu, M.; Zhao, Y.; Zhang, B.; Li, X.; Zhang, L.; Peng, C.; Duan, Y.; Yu, J.; Wang, L.; Yang, K.; Liu, F.; Jiang, R.; Yang, X.; You, T.; Liu, X.; Yang, X.; Bai, F.; Liu, H.; Liu, X.; Guddat, L.W.; Xu, W.; Xiao, G.; Qin, C.; Shi, Z.; Jiang, H.; Rao, Z.; Yang, H.. Structure of Mpro from SARS-CoV-2 and discovery of its inhibitors. Nature. 2020, 582:7811 582, 289–293. <https://doi.org/10.1038/s41586-020-2223-y>.
  95. Mhatre, S.; Srivastava, T.; Naik, S.; Patravale, V. Antiviral activity of green tea and black tea polyphenols in prophylaxis and treatment of COVID-19: A review. Phytomed. 2021, 85, 153286. <https://doi.org/10.1016/J.PHYMED.2020.153286>.
  96. Phong, N.V.; Trang, N.M.; Quyen, C.T.; Anh, H.L.T.; Vinh, L.B. SARS-CoV-2 main protease and papain-like protease inhibition by abietane-type diterpenes isolated from the branches of Glyptostrobus pensilis using molecular docking studies. Nat. Prod. Res. 2022. <https://doi.org/10.1080/14786419.2022.2025801>.
  97. Kılınç, N.; Açar, M.; Tuncay, S.; Karasakal, Ö.F. Identification of potential inhibitors for severe acute respiratory syndrome-related coronavirus 2 (SARS-CoV-2) angiotensin-converting enzyme 2 and the main protease from anatolian traditional plants. Lett. Drug

- Des. Discov. 2022, 19, 996–1006. <https://doi.org/10.2174/1570180819666211230123145>.
98. Nath, M.; Debnath, P. Therapeutic role of traditionally used Indian medicinal plants and spices in combating COVID-19 pandemic situation. *J. Biomol. Struct. Dyn.* 2022, 1–20. <https://doi.org/10.1080/07391102.2022.2093793>.
99. Bahrami, M.; Kamalinejad, M.; Latifi, S.A.; Seif, F.; Dadmehr, M. Cytokine storm in COVID-19 and parthenolide: Preclinical evidence. *Phytother. Res.* 2020, 34, 2429–2430. <https://doi.org/10.1002/PTR.6776>.
100. Park, J.Y.; Jeong, H.J.; Kim, J.H.; Kim, Y.M.; Park, S.J.; Kim, D.; Park, K.H.; Lee, W.S.; Ryu, Y.B. Diarylheptanoids from *alnus japonica* inhibit papain-like protease of severe acute respiratory syndrome coronavirus. *Biol. Pharm. Bull.* 2012, 35, b12-00623. <https://doi.org/10.1248/BPB.B12-00623>.
101. Sinha, S.K.; Shakya, A.; Prasad, S.K.; Singh, S.; Gurav, N.S.; Prasad, R.S.; Gurav, S.S. An in-silico evaluation of different Saikosaponins for their potency against SARS-CoV-2 using NSP15 and fusion spike glycoprotein as targets. *J. Biomol. Struct. Dyn.* 2021, 39, 3244–3255. <https://doi.org/10.1080/07391102.2020.1762741>.